



dasa

**Avaliação da Susceptibilidade aos Glicopeptídeos
em *Staphylococcus* no Laboratório Clínico:
Erros Sutis e Grosseiros**

ALESSANDRO C. O. SILVEIRA

RIF
05

CFO
30

VAN
30

Erro grosseiro

Fenótipos

| Isolate | sVISA | Colony | MIC | | | |
|---------|-------|--------|------|------|------|------|
| | | | 24 h | 48 h | 72 h | 96 h |
| SI4 | no | L | 2 | 3 | 3 | 3 |
| SI11 | yes | P | 2 | 4 | 8 | 12 |
| SI13 | no | L | 2 | 4 | 4 | 4 |
| 10 | yes | P | 1 | 1 | 4 | 16 |
| 36 | no | L | 2 | 2 | 4 | 4 |
| 43 | yes | S | 1 | 2 | 4 | 8 |
| 69 | no | L | 2 | 4 | 4 | 4 |
| 80 | no | L | 2 | 3 | 3 | 4 |

Por que o BRCAST é a melhor opção?

Sem fator de confusão

Com critérios interpretativos para teicoplanina

Sem zona cinza

Com guia epidemiológico de detecção

Considera os testes de triagem para hVISA

| Importância da detecção do mecanismo | |
|---|-----|
| Necessário para categorização clínica da sensibilidade | Sim |
| Para propósito de controle de infecção | Sim |
| Para propósito de saúde pública | Sim |

| Glicopeptídeos ¹ | Ponto de corte p/ CIM (mg/L) | | | | Conteúdo do disco (µg) | Ponto de corte p/ diâmetro do halo (mm) | | | |
|---|------------------------------|---|-----|-----|------------------------|---|-------------------|-------------------|-----|
| | S ≤ | I | R > | AIT | | S ≥ | I | R < | AIT |
| Teicoplanina ² , <i>S. aureus</i> | 2 | - | >2 | | | Nota ^A | Nota ^A | Nota ^A | |
| Teicoplanina, estafilococos coagulase negativo | 4 | - | >4 | | | Nota ^A | Nota ^A | Nota ^A | |
| Vancomicina ² , <i>S. aureus</i> | 2 | - | >2 | | | Nota ^A | Nota ^A | Nota ^A | |
| Vancomicina ² , estafilococos coagulase negativo | 4 | - | >4 | | | Nota ^A | Nota ^A | Nota ^A | |

1. A CIM de glicopeptídeos é dependente do método e deve ser determinada por microdiluição em caldo (referência ISO 20776). *S. aureus* com CIM de 2 mg/L para vancomicina estão no limite da distribuição da CIM do tipo selvagem e pode haver diminuição da resposta clínica. O ponto de corte (resistente) foi diminuído para 2 mg/L para evitar que isolados intermediários "GISA" sejam reportados, já que infecções graves por "GISA" não são tratáveis com doses altas de vancomicina ou teicoplanina.

2. Isolados não sensíveis são raros. A identificação e o teste de sensibilidade em isolados não sensíveis devem ser confirmados em centro de referência.

A. O método de disco-difusão não é confiável e não distingue isolados selvagens daqueles com resistência não mediada pelo gene *vanA*.

hVISA

Resistência heterogênea

Manifesta-se em sub-populações (1 em 10⁶)

Independente da CIM e outros mecanismos

Small Colony Variants (SCV)

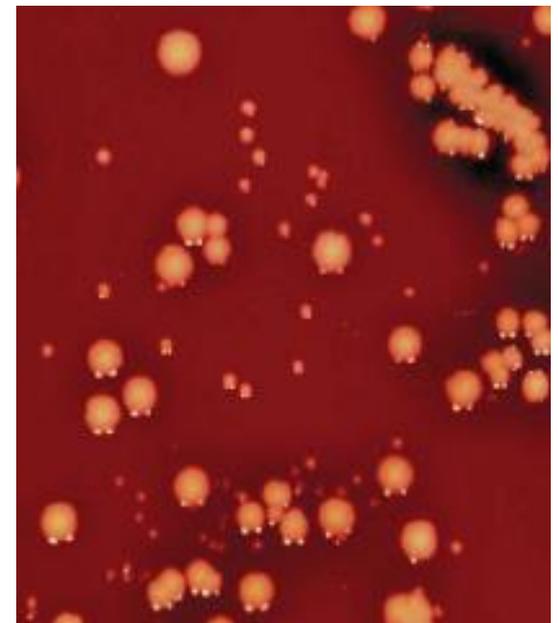
Fator de confusão para o microbiologista

Nutricionalmente exigentes

Crescimento mais lento

Necessidade de pressão seletiva

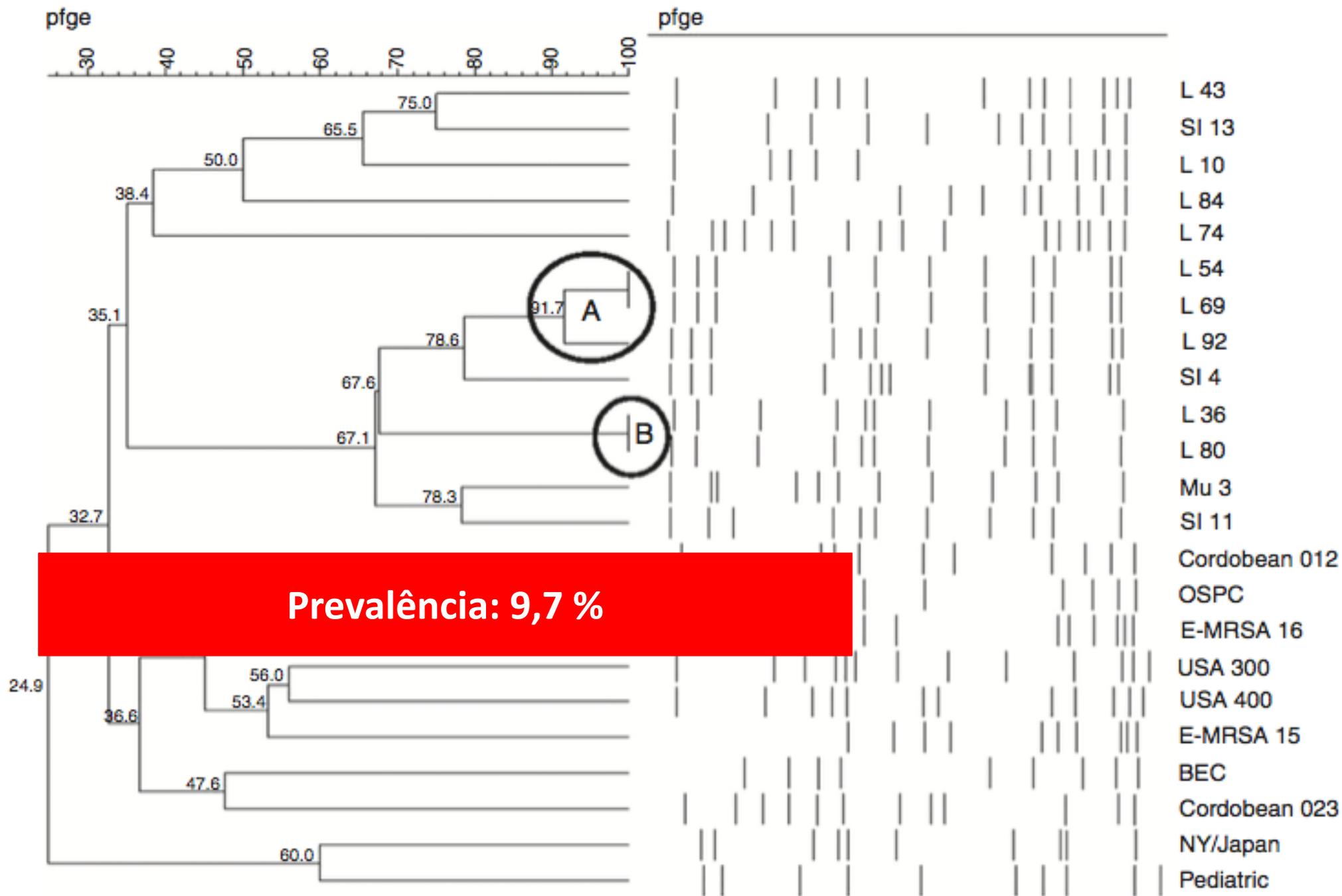
Estágio intermediário para VISA



Systematic Review and Meta-Analysis of the Epidemiology of Vancomycin-Intermediate and Heterogeneous Vancomycin-Intermediate *Staphylococcus aureus* Isolates

Table 2. Prevalence of hVISA and VISA based on study period, origin of study, and isolate selection.^a

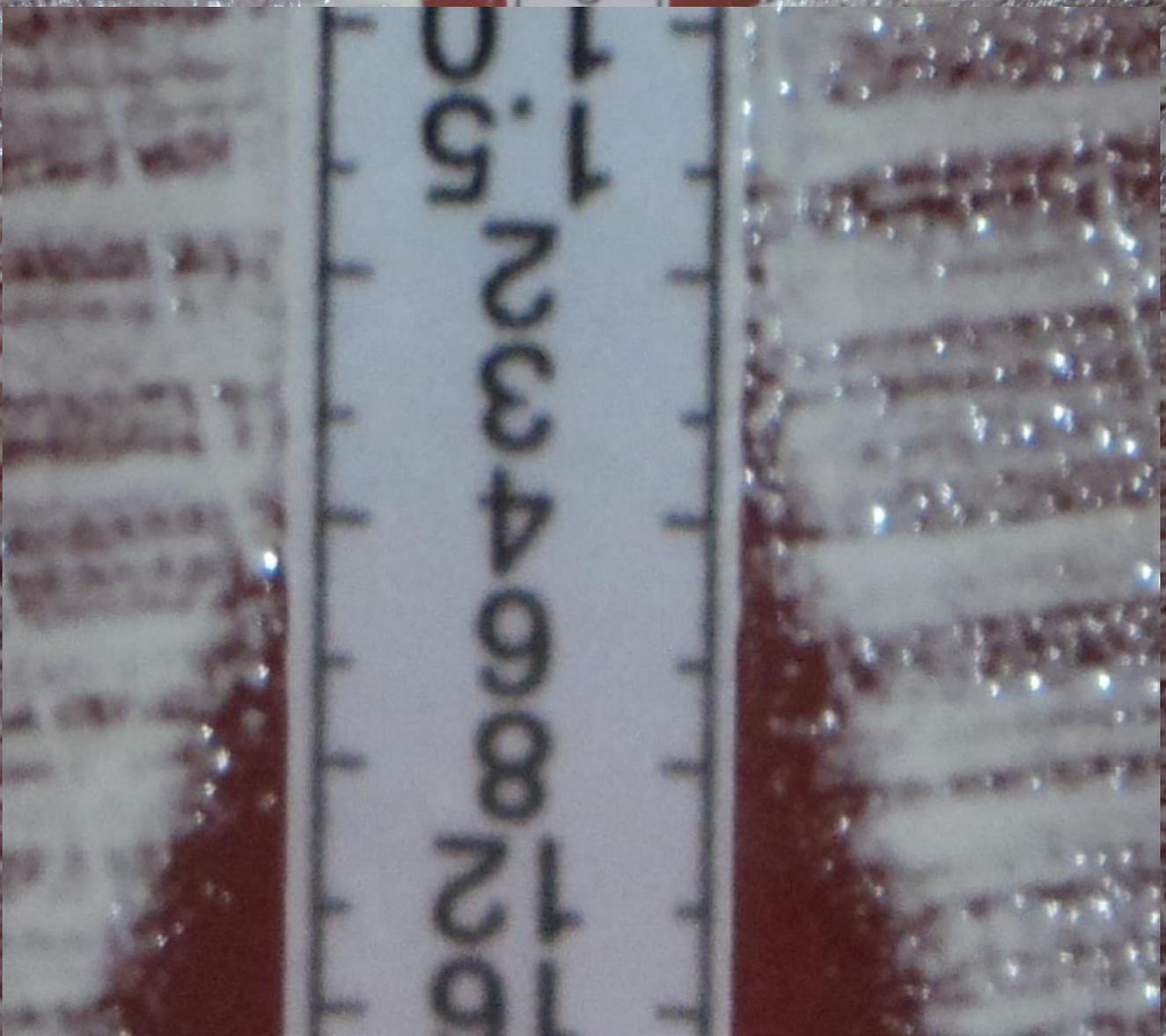
| | Category | Subcategory | No. Studies | No. Strains | Prevalence (%) (95% CI) |
|-------|----------------------|-----------------------|-------------|------------------|-------------------------|
| hVISA | Overall | | 76 | 99042 | 6.05 (4.78–7.48) |
| | Study period | Before 2006 | 42 | 40119 | 4.68 (3.19–6.41) |
| | | 2006–2009 | 10 | 6485 | 5.38 (2.40–9.48) |
| | | 2010–2014 | 5 | 680 | 7.01 (2.12–14.42) |
| | Continent | Asia | 35 | 64692 | 6.81 (4.76–9.16) |
| | | Europe-America | 41 | 34350 | 5.60 (3.85–7.64) |
| | Clinical sample | Blood culture sample | 21 | 5944 | 9.81 (6.71–13.42) |
| | All clinical sample | 55 | 93098 | 4.68 (3.51–6.00) | |
| VISA | Overall | | 38 | 68792 | 3.01 (1.62–4.83) |
| | Study period | Before 2006 | 20 | 13394 | 2.05 (0.95–3.55) |
| | | 2006–2009 | 4 | 5630 | 2.63 (0.29–7.22) |
| | | 2010–2014 | 2 | 2090 | 7.93 (0.06–26.67) |
| | Continent | Asia | 18 | 55362 | 3.42 (1.10–6.99) |
| | | Europe-America | 20 | 13430 | 2.75 (1.19–4.91) |
| | Clinical sample | Blood culture samples | 7 | 2542 | 2.00 (0.03–6.88) |
| | All clinical samples | 31 | 66250 | 3.24 (1.67–5.29) | |



Triagem em ágar com teicoplanina



16
12
8



32
24
16

®
E

VA

256
192
128
96
64
48
32
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16
12
8
6
4
3
2
1.5



Bacteriology

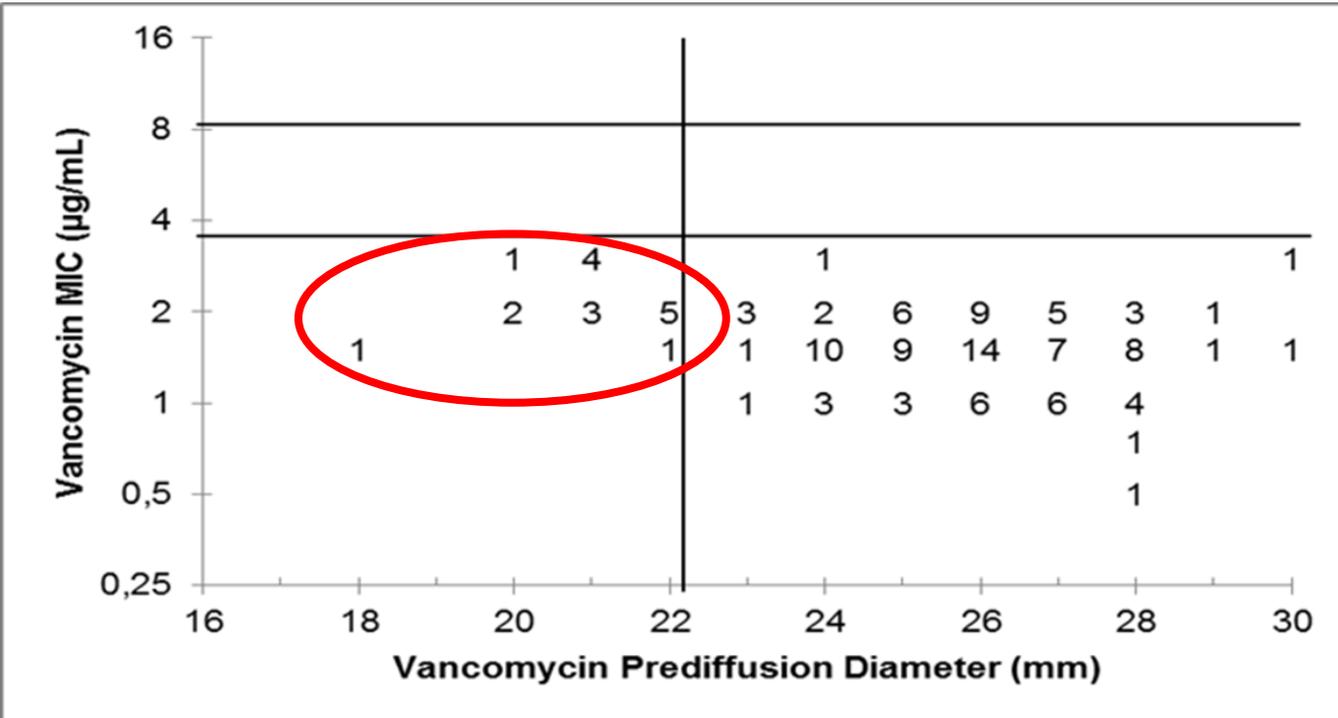
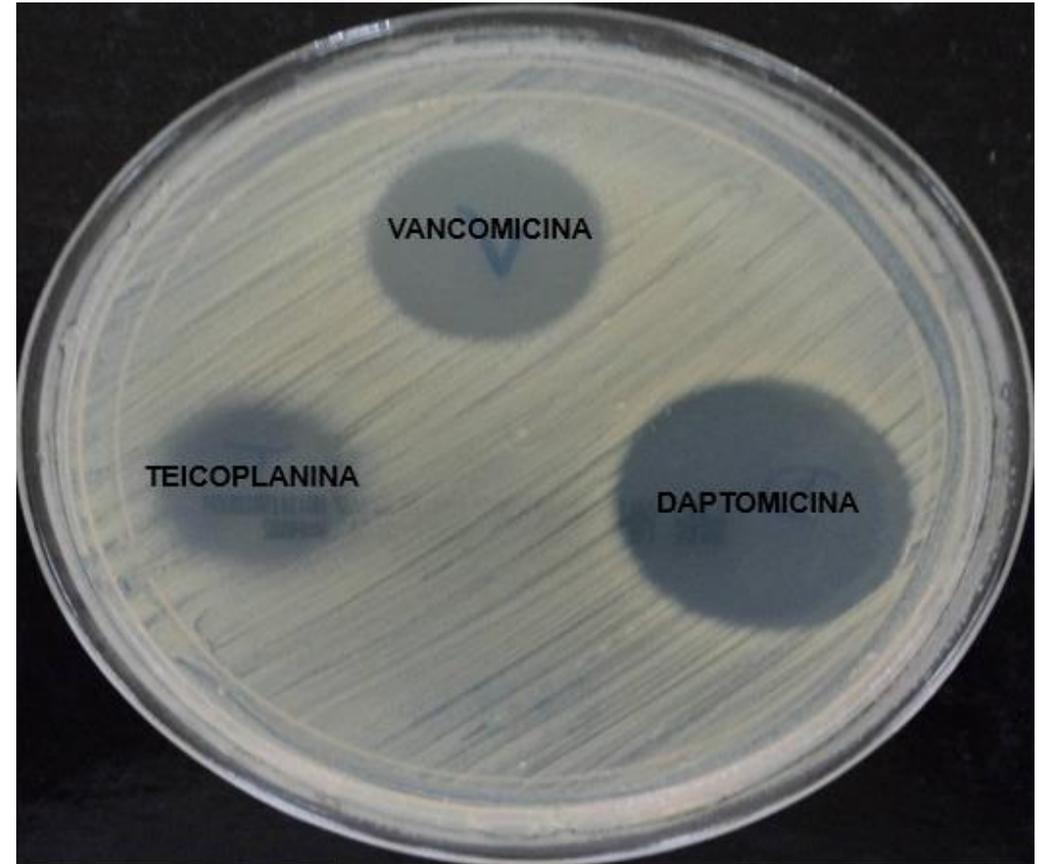
Is prediffusion test an alternative to improve accuracy in screening hVISA strains and to detect susceptibility to glycopeptides/lipopeptides?



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Valor preditivo negativo: 97,1 %

Performance of Various Testing Methodologies for Detection of Heteroresistant Vancomycin-Intermediate *Staphylococcus aureus* in Bloodstream Isolates[▽]

Sebastian J. van Hal,^{1,2†*} Michael C. Wehrhahn,^{1†} Thelma Barbagiannakos,¹ Joanne Mercer,¹ Dehua Chen,¹ David L. Paterson,³ and Iain B. Gosbell^{1,2}

Department of Microbiology & Infectious Diseases, Sydney South West Pathology Service-Liverpool Hospital, Locked Bag 7090, Liverpool BC NSW 1871,¹ Antibiotic Resistance & Mobile Elements Group, School of Medicine, University of Western Sydney, Sydney,² and University of Queensland Centre for Clinical Research (UQCCR), Brisbane,³ Australia

TABLE 2. Comparison of hVISA detection methods to PAP-AUC at hVISA prevalence of 12%^e

| Method (MIC cutoff [$\mu\text{g/ml}$]) | No. of isolates ^a | | Sensitivity (%) | Specificity (%) | NPV (%) | PPV (%) | % Accuracy ^b |
|---|------------------------------|-----------------|-----------------|-----------------|---------|---------|-------------------------|
| | hVISA (TP = 55) | VSSA (TN = 399) | | | | | |
| Etest (≥ 1.5) | 50 | 263 | 91 | 66 | 98 | 27 | 69 |
| BMD (≥ 1.5) | 49 | 335 | 89 | 84 | 98 | 43 | 85 |
| MET | 49 | 218 | 89 | 55 | 97 | 21 | 59 |
| BMD (≥ 2) | 45 | 388 | 82 | 97 | 97 | 80 | 95 |
| Etest (≥ 2) | 39 | 375 | 71 | 94 | 96 | 62 | 91 |
| GRD Etest | 39 | 375 | 71 | 94 | 96 | 62 | 91 |
| Vitek2 (≥ 2) | 14 | 382 | 25 | 96 | 90 | 45 | 87 |

TABLE 3. Sensitivity and specificity values for the different methods in determining hVISA

| Method | Correct no. of hVISA isolates identified (<i>n</i> = 21) | No. of false positives | No. of false negatives | % sensitivity | % specificity |
|---|---|------------------------------|------------------------------|------------------|------------------|
| Etest macromethod, 24 h | 3 | 0 | 18 | 14 | 100 |
| Etest macromethod, 48 h | 12 | 5 | 9 | 57 | 96 |
| Etest GRD, 24 h | 9 | 1 | 12 | 43 | 99 |
| Etest GRD, 48 h | 12 | 3 | 9 | 57 | 97 |
| BHI screen agar, 0.5 McFarland, 24 h | 9 | 0 | 12 | 43 | 100 |
| BHI screen agar, 0.5 McFarland, 48 h | 19 | 8 | 2 | 91 | 94 |
| BHI screen agar, 2 McFarland, 24 h | 16 | 13 | 5 | 76 | 89 |
| BHI screen agar, 2 McFarland, 48 h | 17 | 39 | 0 | 100 | 67 |

Evaluation of the Accuracy of Phenotypic Methods for the Detection of Heteroresistant Vancomycin-Intermediate *Staphylococcus aureus* (hVISA)

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²Department of Pharmaceutical Sciences, Regional University of Blumenau, Brazil

JSM Microbiology 4(1): 1031 (2016)

Table 2: Parameters of the main screening tests for the detection of hVISA.

| Methodology | Sensitivity | Specificity | PPV ^a | NPV ^b | Accuracy |
|-----------------------------|-------------|-------------|------------------|------------------|----------|
| Etest GRD ^c | 66.7% | 97.3% | 72.3% | 96.5% | 94.3% |
| Etestmacromethod | 75% | 94.6% | 60% | 97.2% | 92.7% |
| Agar screening ^d | 90.9% | 93.8% | 58.8% | 99.1% | 93.5% |

^a– Positive predictive value

^b– Negative predictive value

^c– Etest glycopeptides resistance detection[®]

^d– Agar screening in brain-heart infusion (BHI) with 4 µg/mL vancomycin and 16 g/L pancreatic digest of casein

Erro sutil

Rapid Detection of Vancomycin-Intermediate *Staphylococcus aureus* by Matrix-Assisted Laser Desorption Ionization–Time of Flight Mass Spectrometry

Cheryl A. Mather,^{a,d*} Brian J. Werth,^b Shobini Sivagnanam,^c Dhruva J. SenGupta,^a Susan M. Butler-Wu^{a*}

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Rapid and easy detection of low-level resistance to vancomycin in methicillin-resistant *Staphylococcus aureus* by matrix-assisted laser desorption ionization time-of-flight mass spectrometry

Kota Asakura¹, Takuya Azechi¹, Hiroshi Sasano¹, Hidehito Matsu², Hideaki Hanaki², Motoyasu Miyazaki³, Tohru Takata⁴, Miwa Sekine⁵, Tomoiku Takaku⁶, Tomonori Ochiai⁶, Norio Komatsu⁶, Keigo Shibayama^{7,8}, Yuki Katayama^{5e*}, Koji Yahara^{8e*}

PLOS ONE | <https://doi.org/10.1371/journal.pone.0194212> March 9, 2018

← → ↻ 127.0.0.1:8888

hVISA Classifier

Path to a directory of data for prediction

Path to a .RData file of spectra data

or to a directory of raw spectra data

Cross validation

Analyze

Path to an output CSV to be displayed

Display CSV

hVISA Classifier

Path to a directory of data for prediction

Path to a .RData file of spectra data

or to a directory of raw spectra data

Cross validation

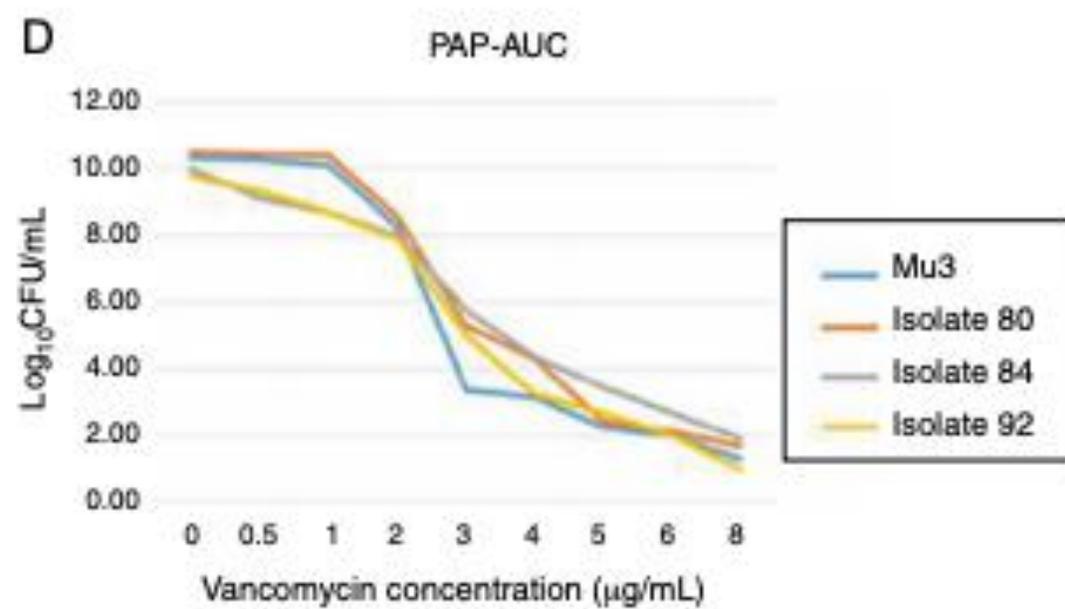
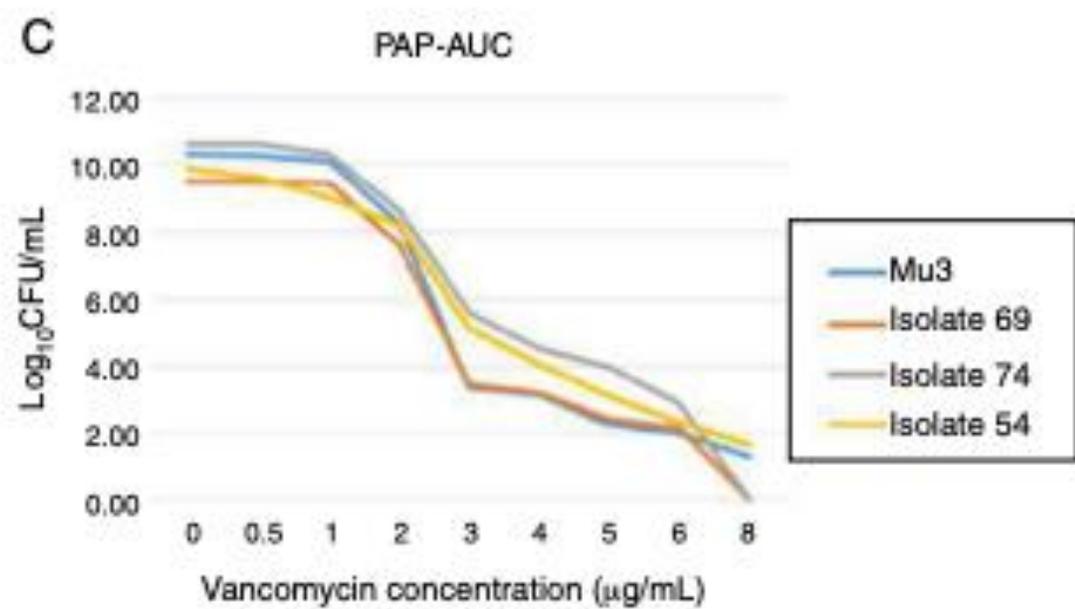
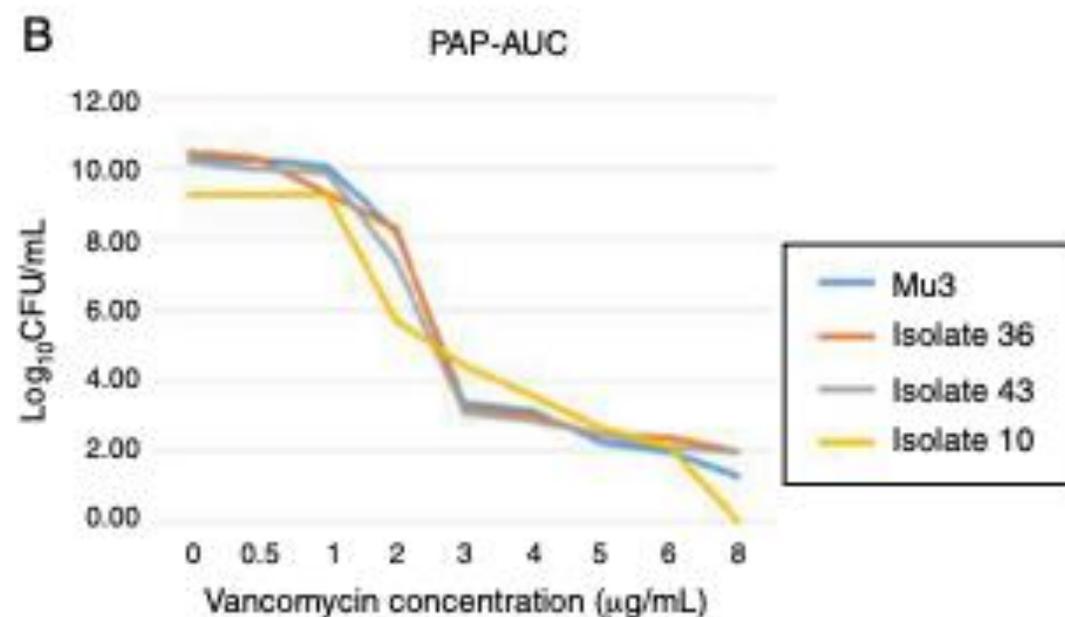
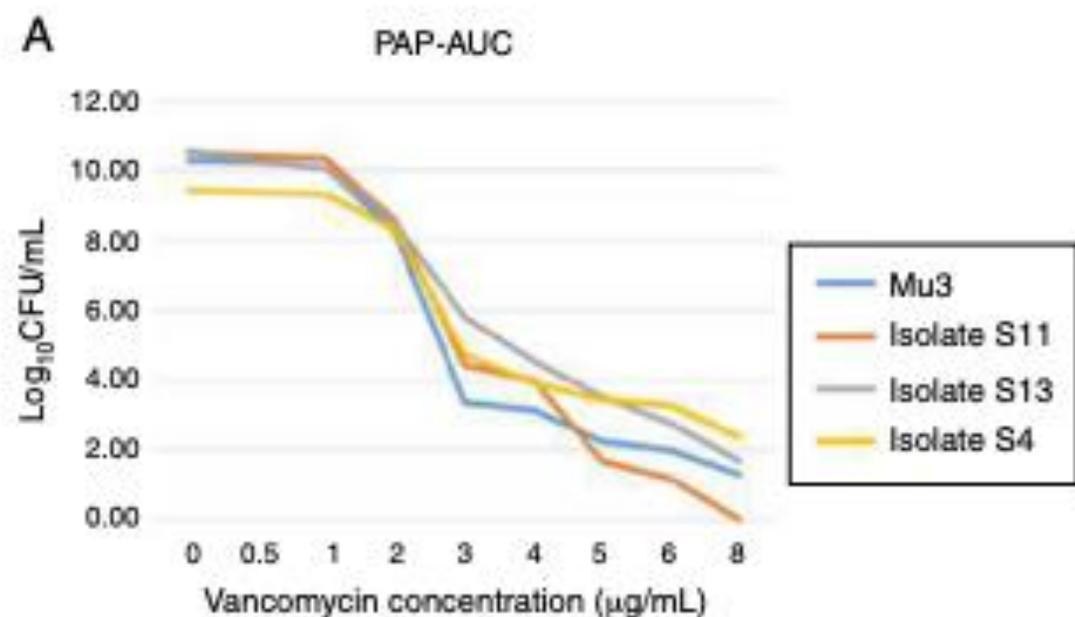
Analyze

Path to an output CSV to be displayed

Display CSV

create CSV file -> prediction_of_example_data_out.csv

| sample | prediction |
|--|------------|
| C:\hVISAclassifier\example_data\hVISA\0916-MRSA5\0_B10\1\1SLin\fid | hVISA |
| C:\hVISAclassifier\example_data\hVISA\9_hVISA_MRSA5\0_E6\1\1SLin\fid | hVISA |
| C:\hVISAclassifier\example_data\hVISA\MRSA239-3\0_F10\1\1SLin\fid | hVISA |
| C:\hVISAclassifier\example_data\hVISA\MRSA81-3\0_D6\1\1SLin\fid | hVISA |
| C:\hVISAclassifier\example_data\VISA\MRSA372\0_D11\1\1SLin\fid | VISA |
| C:\hVISAclassifier\example_data\VISA\T11\0_B3\2\1SLin\fid | VISA |
| C:\hVISAclassifier\example_data\VSSA\FU_97\0_F11\1\1SLin\fid | VSSA |
| C:\hVISAclassifier\example_data\VSSA\VSSA104\0_C1\1\1SLin\fid | VSSA |



Clinical and Microbiological Characteristics of Heteroresistant and Vancomycin-Intermediate *Staphylococcus aureus* from Bloodstream Infections in a Brazilian Teaching Hospital

PLOS ONE | DOI:10.1371/journal.pone.0160506 August 30, 2016

Thaina Miranda da Costa¹, Priscylla Guimarães Miguereles Morgado¹, Fernanda Sampaio Cavalcante¹, Andreia Paredes Damasco¹, Simone Aranha Nouér², Kátia Regina Netto dos Santos^{1*}

Table 4. Microbiological characteristics of six *Staphylococcus aureus* isolates presenting vancomycin MIC of 2 mg/L with at least one screening test positive to detect heterogeneous vancomycin intermediate resistance (hVISA).

| Isolatenumber | Methicillin resistance/SCCmec type | Broth microdilution test MIC (mg/L) ^a | | | Screening plates (48h of incubation) | | | | Etest macro ^b (µg/mL) | | Etest GRD ^b (µg/mL) | | PAP-AUC ratio ^c | Interpretation (Clonality) |
|---------------|---|--|------|-----|--------------------------------------|-------|--------|-------|----------------------------------|-----|--------------------------------|-----|----------------------------|----------------------------|
| | | TEI | OXA | DAP | BHIa3 | BHIa4 | BHI4ca | BHIa6 | VAN | TEI | VAN | TEI | | |
| 1594 |  SCCmecII | 0.5 | 128 | 2 | + | + | + | - | 3 | 16 | 1 | 3 | 1.15 | hVISA (USA100/ST5/CC5) |
| 1636 | SCCmecIV | 0.25 | 8 | 1 | + | - | - | - | 4 | 4 | 1 | 1.5 | 0.50 | False positive |
| 1588 | MSSA | 0.25 | 0.5 | 1 | - | - | - | - | 4 | 12 | 1 | 3 | ND | NA |
| 1595 | MSSA | 0.5 | 0.5 | 2 | - | - | - | - | 3 | 16 | 0.5 | 3 | ND | NA |
| 1622 | MSSA | 0.25 | 0.25 | 1 | + | - | - | - | 2 | 2 | 0.75 | 1.5 | ND | NA |
| 1691 | MSSA | 0.25 | ≤0.2 | 1 | + | - | - | - | 6 | 4 | 0.75 | 3 | ND | NA |

Infecção por MRSA

**Hemocultura
Abscesso
Trato respiratório inferior**

**CIM Vancomicina $\geq 1,5$ mg/L (Etest) ou
2 mg/L (microdiluição)**

Triagem para hVISA

**Ágar BHI com 5 mg/L de teicoplanina
Inóculo de 2 na escala de McFarland
Incubação por 48 horas
Spot de 10 ul**

Crescimento de 2 ou mais colônias

**Sugerir modificação de terapia
com vancomicina**

**Encaminhar microrganismo para
laboratório de referência**

Mortalidade

Systematic Review and Meta-Analysis of the Significance of Heterogeneous Vancomycin-Intermediate *Staphylococcus aureus* Isolates[▽]

Sebastiaan J. van Hal^{1*} and David L. Paterson²

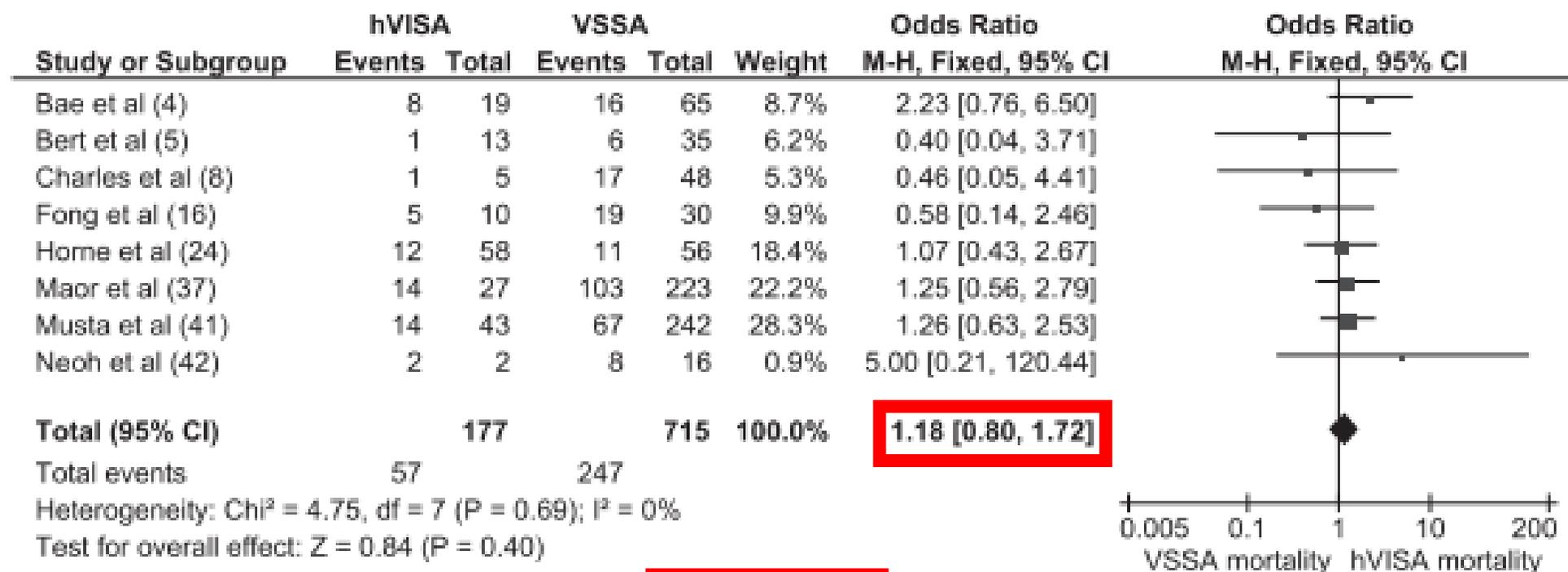


FIG. 2. Forest plot (using Mantel-Haenszel analysis) of 30-day mortality in hVISA- compared to VSSA-infected patients with “events” denoting deaths in each group. Squares indicate point estimates, and the size of the square indicates the weight of each study.

Mortalidade

Table 3. Clinical characteristics of the six patients with bloodstream infections caused by vancomycin intermediate *Staphylococcus aureus* (VISA) isolates.

| Isolate number | Cause of admission | Gender | Charlson score | Dialysis | Ward | Previous therapy | Treatment after bacterial isolation | Outcome (number of days from therapy to outcome); Type of VISA infection |
|----------------|---|--------|----------------|----------------------|-------------------|--|--|--|
| 1579 | Community-acquired Pneumonia | M | 6 | Acute hemodialysis | ICU | amx + sul + azm// tzp // tei + tzp //tei //van + mem + flu | van + mem + flu | Death (6); VAP infection  |
| 1582 | Bacteremia (VISA) due to dialysis | F | 4 | Peritoneal dialysis | Nephrology | Amk | van + amk//van | Discharge (13); Bacteremia related to dialysis |
| 1616 | Liver transplantation | F | 4 | Acute hemodialysis | ICU | nor + cro// sul + flu + amx// van + pmb + mem | Van | Death (4); Bacteremia due to central vascular catheter  |
| 1698 | Infection (VISA) after vascular surgery | M | 3 | No | Internal Medicine | van // sxt // fep// tei | tei // dap // sxt + dap // gen + dap// dap // dap + fep // cip | Discharge (59); Nosocomial osteomyelitis with endocarditis |
| 1638 | Cancer | F | 6 | No | Oncology | fep// fep + van | fep // fep + van // oxa | Discharge (36); Bacteremia due to peripheral vascular catheter |
| 1645 | Bacteremia due to hemodialysis | M | 2 | Chronic hemodialysis | Nephrology | Cip | cfz // oxa + amk// cfz | Discharge (16); Bacteremia due to central vascular catheter |

Perspectivas Futuras

Realização da microdiluição em caldo

Estudos clínicos para avaliar impacto na mortalidade

Programa de detecção e monitoramento da resistência, em parceria com o CGLAB

Epidemiologia molecular de MRSA no Brasil

Validação da pré-difusão

Estudo do MALDI-TOF com amostras brasileiras

Utilização de outras alternativas terapêuticas: tigeciclina, daptomicina, linezolida, clindamicina, sulfametoxazol-trimetoprim

Uso de marcador molecular

Conclusões

A detecção da diminuição da susceptibilidade à vancomicina é um grande desafio

Negligenciada no Brasil

Cara → elitizada

Problema grave de saúde pública

Associada à morbidade e mortalidade

Imprescindível a criação de programas públicos para detecção, controle e monitoramento

Fundamental para diminuição de mortalidade e custos em saúde

Obrigado!

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